Human Peroxisome proliferator-activated receptor gamma coactivator 1 alpha ELISA KIT

BIOENZY
Rev.04-2

Catalog no	Size
BZ-08129053-EA	48 Wells
BZ-08129053-EB	96 Wells

User Instruction

Standard Curve Range	0.05 ng/ml – 30 ng/ml
Sensitivity	0.021 ng/ml
Repeatability	Intra-Assay: CV < 8%; Inter-Assay: CV < 10%
Storage	Keep the **whole kit** before use in: • -20°C for 12 months validity • 2-8°C for 6 months validity Avoid repeated freeze–thaw cycles. Do not use the kit beyond its expiration date.

Specificity

This Sandwich kit is for the accurate quantitative detection of Human Peroxisome Proliferator-Activated Receptor Gamma Coactivator 1 Alpha (also known as PPARGC1A) in serum, plasma, cell culture supernatants, Ascites, tissue homogenates or other biological fluids.

Assay Principle

This kit is an Enzyme-Linked Immunosorbent Assay (ELISA). The plate has been pre-coated with Human PGC1α antibody. PGC1α present in the sample is added and binds to antibodies coated on the wells. And then biotinylated Human PGC1α Antibody is added and binds to PGC1α in the sample. Then Streptavidin-HRP is added and binds to the Biotinylated PGC1α antibody. After incubation unbound Streptavidin-HRP is washed away during a washing step. Substrate solution is then added and color develops in proportion to the amount of Human PGC1α. The reaction is terminated by addition of acidic stop solution and absorbance is measured at 450 nm.

Storage Directions and Kit Components

Keep the **whole kit** before use in:

- -20°C for 12 months validity
- 2-8°C for 6 months validity

After opening handling, store the items separately according to the following directions:

Components	96 Wells	48 Wells	Storage Directions After Opening Handling
Pre-Coated Microplate	12 x 8 wells	12 x 4 wells	2-8°C (1 week) / -20°C (1 months)
Standard Solution (32 ng/ml)	0.5 ml	0.5 ml	
Biotinylated Human PGC1α Antibody	1 ml	1 ml	
Streptavidin-HRP	6 ml	3 ml	
Standard Diluent	3 ml	3 ml	
Stop Solution	6 ml	3 ml	
Wash Buffer Concentrate (25x)	20 ml	20 ml	
Substrate Solution A	6 ml	3 ml	
Substrate Solution B	6 ml	3 ml	2-8°C (1 week) / -20°C (1 months) (light sensitive, avoid light)
Plate Covers	2	2	RT
Instruction manual	1	1	

Notes:

- If the whole kit is stored at -20°C, place the kit at 2-8°C the day before the experiment.
- After receiving the kit, the plates can be disassembled to single strips. Please use it in batches on demands.

Material Required But Not Supplied

- 37°C±0.5°C incubator
- Absorbent paper
- Precision pipettes and disposable pipette tips
- Clean tubes
- Deionized or distilled water
- Microplate reader with 450 ± 10nm wavelength filter

Precautions

- Prior to use, the kit and sample should be warmed naturally to room temperature 30 minutes.
- This instruction must be strictly followed in the experiment.

- Once the desired number of strips has been removed, immediately reseal the bag to protect the remaining from deterioration. Cover all reagents when not in use.
- Make sure pipetting order and rate of addition from well-to-well when pipetting reagents.
- Pipette tips and plate sealer in hand should be clean and disposable to avoid cross-contamination.
- Avoid using the reagents from different batches together.
- Substrate solution B is sensitive to light, don't expose substrate solution B to light for a long time.
- Stop solution contains acid. Please wear eye, hand and skin protection when using this material. Avoid contact of skin or mucous membranes with kit reagent.
- The kit should not be used beyond the expiration date.

Specimen Collection

Serum Allow serum to clot for 10-20 minutes at room temperature. Centrifuge at 2000-3000 RPM for 20 minutes. Collect the supernatant without sediment.

Plasma Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 20 minutes at 2000-3000 RPM within 10-20 minutes of collection. Collect the supernatant without sediment.

Urine/Ascites/Cerebrospinal fluid Collect by sterile tube. Centrifuge at 2000-3000 RPM for approximately 20 minutes. Collect the supernatant without sediment.

Cell Culture Supernatant Collect by sterile tubes when examining secrete components. Centrifuge at 2000-3000 RPM for approximately 20 minutes. Collect the supernatants carefully. When examining the components within the cell, use PBS (pH 7.2-7.4) to dilute cell suspension to the cell concentration of approximately 1 million/ml. Damage cells through repeated freeze-thaw cycles to let out the inside components. Centrifuge at 2000-3000 RPM for approximately 20 minutes. Collect the supernatant without sediment.

Tissue Rinse tissues in ice-cold PBS (pH 7.4) to remove excess blood thoroughly and weigh before homogenization. Mince tissues and homogenize them in PBS (tissue weight (g): PBS (mL) volume=1:9) with a glass homogenizer on ice. To further break down the cells, you can sonicate the suspension with an ultrasonic cell disrupter or subject it to freeze-thaw cycles. The homogenates are then centrifuged for 15 minutes at 12,000 RPM at 4℃ to get the supernatant.

Note

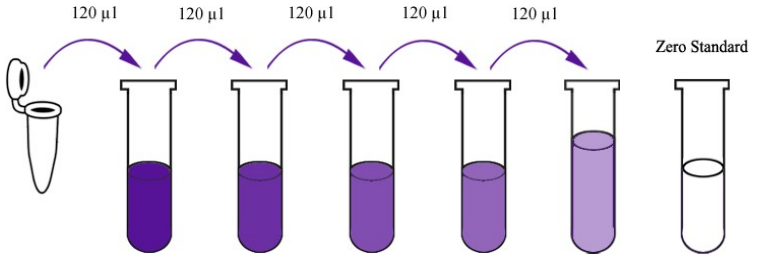
- Sample concentrations should be predicted before being used in the assay. If the sample concentration is not within the range of the standard curve, users must **contact us** to determine the optimal sample for their particular experiments.
- Samples to be used within 5 days should be stored at 2-8℃. Samples should be aliquoted or must be stored at -20℃ within 1 month or -80℃ within 3 months. Avoid repeated freeze thaw cycles.
- Samples should be brought to room temperature before starting the assay.
- Centrifuge to collect sample before use.
- Samples containing NaN3 can't be tested as it inhibits the activity of Horse Radish Peroxidase (HRP).
- Collect the supernatants carefully. When sediments occurred during storage, centrifugation should be performed again.
- Hemolysis can greatly impact the validity of test results. Take care to minimize hemolysis.

**Sample can't be diluted with this kit. Owing to the material we use to prepare the kit, the sample matrix interference may falsely depress the specificity and accuracy of the assay.*

Reagent Preparation

- All reagents should be brought to room temperature before use.
- **Standard** Reconstitute the 120 µl of the standard (32 ng/ml) with 120 µl of standard diluent to generate a 16 ng/ml standard stock solution. Allow the standard to sit for 15 mins with gentle agitation prior to making dilutions. Prepare duplicate standard points by serially diluting the standard stock solution (320 ng/ml) 1:2 with standard diluent to produce 160 ng/ml, 80 ng/ml, 40 ng/ml, and 20 ng/ml solutions. Standard diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20℃ and used within one month. Dilution of standard solutions suggested are as follows:

16 ng/ml	Standard No.5	120 µl Original Standard + 120 µl Standard Diluent
8 ng/ml	Standard No.4	120 µl Standard No.5 + 120 µl Standard Diluent
4 ng/ml	Standard No.3	120 µl Standard No.4 + 120 µl Standard Diluent
2 ng/ml	Standard No.2	120 µl Standard No.3 + 120 µl Standard Diluent
1 ng/ml	Standard No.1	120 µl Standard No.2 + 120 µl Standard Diluent



Standard Concentration	Standard No.5	Standard No.4	Standard No.3	Standard No.2	Standard No.1
32 ng/ml	16 ng/ml	8 ng/ml	4 ng/ml	2 ng/ml	1 ng/ml

- **Wash Buffer** Dilute 20 ml of Wash Buffer Concentrate 25x into deionized or distilled water to yield 500 ml of 1x Wash Buffer. If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.

Assay Procedure

1. Prepare all reagents, standard solutions and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature.
2. Determine the number of strips required for the assay. Insert the strips in the frames for use.

The unused strips should be stored at 2-8℃.

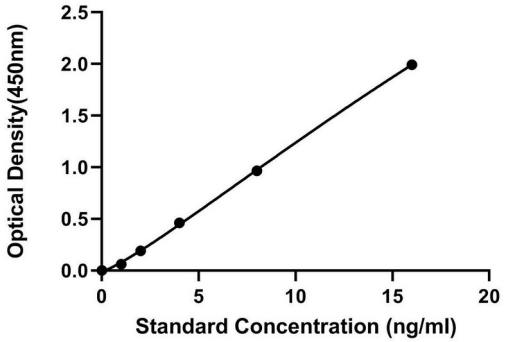
3. Add 50 µl standard to standard well. **Note**: Don't add biotinylated antibody to standard well because the standard solution contains biotinylated antibody.
4. Add 40 µl sample to sample wells and then add 10 µl anti-PGC1α antibody to sample wells, then add 50 µl streptavidin-HRP to sample wells and standard wells (Not blank control well). Mix well. Cover the plate with a sealer. Incubate 60 minutes at 37℃.
5. Remove the sealer and wash the plate 5 times with wash buffer. Soak wells with 300 µl wash buffer for 30 seconds to 1 minute for each wash. For automated washing, aspirate all wells and wash 5 times with wash buffer, overfilling wells with wash buffer. Blot the plate onto paper towels or other absorbent material.
6. Add 50 µl substrate solution A to each well and then add 50 µl substrate solution B to each well. Incubate plate covered with a new sealer for 10 minutes at 37℃ in the dark.
7. Add 50 µl Stop Solution to each well, the blue color will change into yellow immediately.
8. Determine the optical density (OD value) of each well immediately using a microplate reader set to 450 nm within 10 minutes after adding the stop solution.

Calculation of Result

Construct a standard curve by plotting the average OD for each standard on the vertical (Y) axis against the concentration on the horizontal (X) axis and draw a best fit curve through the points on the graph. These calculations can be best performed with computer-based curve-fitting software and the best fit line can be determined by regression analysis.

Typical Data

This standard curve is only for demonstration purposes. A standard curve should be generated with each assay.



Summary

1. Prepare all reagents, samples and standards.
2. Add sample and ELISA reagent into each well. Incubate for 1 hour at 37℃.
3. Wash the plate 5 times.
4. Add substrate solution A and B. Incubate for 10 minutes at 37℃.
5. Add stop solution and color develops.
6. Read the OD value within 10 minutes.

Shipping Direction - Temperature

Maintain temperature between 2℃ to 8℃ during transportation in unopened packaging.

Troubleshooting

Possible Case	Solution
High Background	
• Improper washing • Substrate was contaminated • Non-specific binding of antibody • Plate are not be sealing incompletely • Incorrect incubation temperature • Substrate exposed to light prior to use • Contaminated wash buffer	• Increasing duration of soaking steps • Replace. Substrate should be clean and avoid crossed contamination by using the sealer • Replace another purified antibody or blocking buffer • Make sure to follow the instruction strictly • Incubate at room temperature • Keep substrate in a dark place • Use a clean buffers and sterile filter
Weak Signal	
• Improper washing • Incorrect incubation temperature • Antibody are not enough • Reagent are contaminated • Pipette are not clean	• Increasing duration of soaking steps • Incubate at room temperature • Increase the concentration of the antibody • Use new one • Pipette should be clean
No Signal	
• Reagent are contaminated • Sample prepared incorrectly • Antibody are not enough • Wash buffer contains sodium azide • HRP was not added	• Use new one • Make sure the sample workable/dilution • Increase the antibody concentration • Use a new wash buffer and avoid sodium azide in it • Add HRP according to the instruction
Poor Precision	
• Imprecise/ inaccurate pipetting • Incomplete washing of the wells	• Check/ calibrate pipettes Make sure wells are washed adequately by filling the wells with wash buffer and all residual antibody solutions crossed well before washing.

This product is for research use only, not for use in diagnosis procedures.
It's highly recommended to read this instruction entirely before use.

For more information:
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